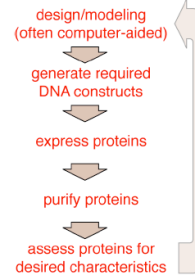


System Engineering

20.109(F10)
M2D6 lecture
11.02.10

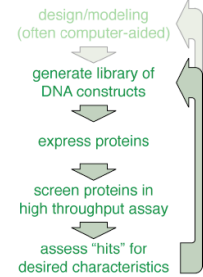
Rational protein design:

Knowledge-based, deterministic engineering of proteins with novel characteristics



“Irrational” high throughput protein engineering:

Selection for desired properties from libraries of random variants



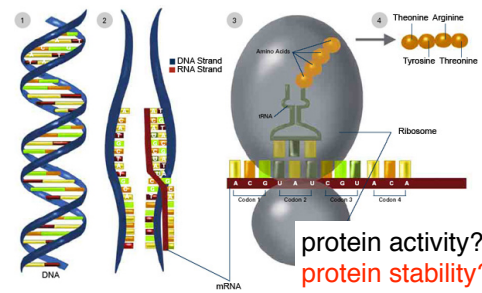
Slide from Alan Jasanoff

Design: rational vs “irrational”

Cph1/EnvZ	A553	G554	V555	S556	H557
EnvZ	A239T	G240E	V241G	S242D	H243A
wt seq	GCG	GGG	GTA	AGT	CAC
oligo seq	RNS	RNS	RNS	RNS	SNW
poss aa	Val	Val	Val	Val	Val
	Ala	Ala	Ala	Ala	Ala
	Asp	Asp	Asp	Asp	Asp
	Glu	Glu	Glu	Glu	Glu
	Gly	Gly	Gly	Gly	Gly
	Ile	Ile	Ile	Ile	Leu
	Met	Met	Met	Met	Pro
	Thr	Thr	Thr	Thr	His
	Asn	Asn	Asn	Asn	Gln
	Lys	Lys	Lys	Lys	Arg
	Ser	Ser	Ser	Ser	
	Arg	Arg	Arg	Arg	

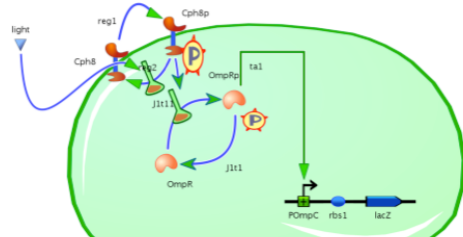
NOTE: no stop codons should be in mix

Beyond the C-dog



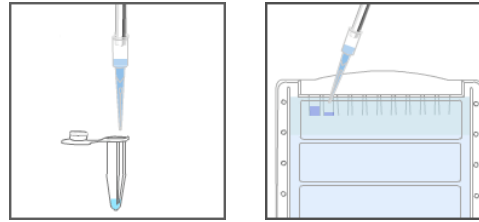
http://publications.nigms.nih.gov/thenewgenetics/images/ch1_trans.jpg

How could a change in protein stability affect β -gal?



	# autophos Cph8	Kinasing rate	P-tase rate	Fraction OmpR+P
Wild type	10	1	0.5	1 of 10

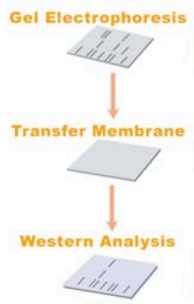
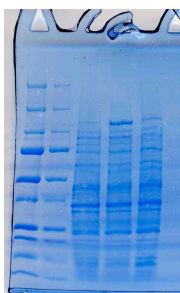
Part 1: SDS-Polyacrylamide Gel Electrophoresis (SDS-PAGE)



Loading dye has glycerol, SDS, reducing agent

Samples boiled before loading

Part 2: Transfer for Western



Thursday:
probe
membrane
with an
antibody

http://www.genscript.com/product_001/western_application/grp_id/60065/op/detail/Uvrag_Antibody_Analysis.html

20.109(F10): Laboratory Fundamentals of Biological Engineering

In lab you will:

- Measure OD600 of 1:10 of bacterial photography strain, Candidate 1, Candidate 2
- Harvest 4 OD
e.g. if 0.5 OD, harvest 8 ml of 1:10 or 0.8 ml of undiluted
- Isolate protein with lysis kit (enzymatic lysis of cells, spin out debris)
- Mix supernatant with loading dye
- Boil
- Load for SDS-PAGE along with markers, + control lysate

Order#	Req#	SeqId	Download	View	Sample	Primer	UID	Date	Phred Q20	Comments
31477	379712	4491-1	Text Chromat	View	Red-334	NO289	1742	Oct 28 2010	0	being repeated-failed
31477	379713	4491-2	Text Chromat	View	Red-cand1	NO289	1742	Oct 28 2010	rhd qual 189 fasta	being repeated-noisy/failed
31477	379714	4491-3	Text Chromat	View	Red-cand2	NO289	1742	Oct 28 2010	rhd qual 17 fasta	being repeated-noisy/failed
31477	379715	4491-4	Text Chromat	View	Orange-334	NO289	1742	Oct 28 2010	0	being repeated-failed
31477	379716	4491-5	Text Chromat	View	Orange-cand1	NO289	1742	Oct 28 2010	0	being repeated-failed
31477	379717	4491-6	Text Chromat	View	Orange-cand2	NO289	1742	Oct 28 2010	rhd qual 812 fasta	Results Available
31477	379718	4491-7	Text Chromat	View	Yellow-334	NO289	1742	Oct 28 2010	rhd qual 1000 fasta	Results Available
31477	379719	4491-8	Text Chromat	View	Yellow-cand1	NO289	1742	Oct 28 2010	rhd qual 946 fasta	Results Available
31477	379720	4491-9	Text Chromat	View	Yellow-cand2	NO289	1742	Oct 28 2010	0	being repeated-failed
31477	379721	4491-10	Text Chromat	View	Green-334	NO289	1742	Oct 28 2010	rhd qual 987 fasta	Results Available
31477	379722	4491-11	Text Chromat	View	Green-cand1	NO289	1742	Oct 28 2010	0	being repeated-failed
31477	379723	4491-12	Text Chromat	View	Green-cand2	NO289	1742	Oct 28 2010	rhd qual 12 fasta	being repeated-noisy/failed
31477	379724	4491-13	Text Chromat	View	Blue-334	NO289	1742	Oct 28 2010	rhd qual 992 fasta	Results Available
31477	379725	4491-14	Text Chromat	View	Blue-cand1	NO289	1742	Oct 28 2010	rhd qual 922 fasta	Results Available
31477	379726	4491-15	Text Chromat	View	Blue-cand2	NO289	1742	Oct 28 2010	rhd qual 104 fasta	being repeated-noisy/failed

First place to look: your notebook!

6-4-76 UV Spectrum of Guanine from Adduct

Introduction: The experiment on p. 17 failed to produce an interpretable UV spectrum. The Gua peak was reaspirated to dryness as described above and dissolved in H₂O (100µl) as described above.

- Inject into HPLC eluted under TRN+ conditions
- Collect peak in Cuvette
- Obtain UV spectrum

Chromatographic Results:

1. Gua from adduct : 95% on X.1 (254 nm)
2. Gua standard : 60% on X.2

Spectroscopic Results : facing page.

Conclusions: The adduct contains a base with an identical UV spectrum to guanine.

Today in lab

1. Lab treat
2. SDS-PAGE + blot
3. β -gal assay for cells grown in light and dark
4. Analysis of sequence data

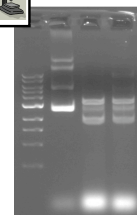
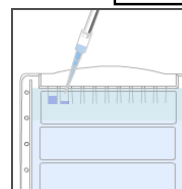
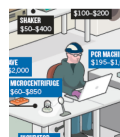
BEFORE LEAVING LAB today:

5' with NK, NT and/or MS to discuss what you and your partner would like to do...

Consider repeating experiments that need to be repeated

Consider collaborating with other groups to get more interpretable/robust data

Summary



20.109(F10): Laboratory Fundamentals of Biological Engineering